

COCONUT PALM DISEASES CAUSED BY FUNGI, BACTERIA, PHYTOPLASMA AND VIRUS – AN ULTIMATUM TO PALMS OF GLOBAL ECONOMY

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Abstract

Coconut is a predominant crop cultivated in more than 93 countries in the world, especially in tropical regions. Among the major coconut cultivation states in India such as Kerala, Karnataka, Odisha, Maharashtra, Tamil Nadu, Gujarat, Assam and Andra Pradesh, Kerala has the longest history of cultivation. However, it is affected by various kinds of diseases such as lethal yellowing, red-ring, bud rot, ganodarma, stem bleeding, leaf rot, leaf blight, root wilt and cadang-cadang diseases leading significant losses in production. Major coconut products in the market includes Kernal Based, Water based, Coconut shell based, inflorescence based and convenience based items. It consists of virgin coconut oil, neera, coconut milk, desiccated coconut powder, activated carbon, shell powder, ball copra, tender coconut water, coconut vinegar, coconut wood, shell charcoal and coconut handicrafts. This paper reviews the common fungi, bacteria, phytoplasma and virus diseases found in coconut and their symptoms. The review concludes that proper management strategies should be taken for reducing the impact of diseases on coconut and thereby maintaining the productivity.

INTRODUCTION

On the edge of escalating popularity of coconut among health-conscious customers, countries are implementing various strategies to increase their production in coconut. Coconut farming plays a salient part in the agrarian economy of Kerala and ranked one in the production of Coconut besides its unique place ' in the socio-cultural fabric of the region. Coconut has been cultivated in large number of countries, around more than 93 countries in the world having a high economic and social impact and India place one of the top among them. Coconut production in Kerala contributes major part in the economy [1]. According to the world wide census conducted by Food and Agriculture Organization from the years 2016 to 2020, top 10 countries producing coconut are India, Indonesia, Philippines, Brazil, Sri Lanka, Vietnam, Papua New guinea, Mexico, Thailand and Malaysia [2]

Rainfall during 3rd to 4th quarter has negative impact on coconut production whereas rainfall on first and second quarter has positive impact on the production of coconut. Coconut cultivation areas are shrinking drastically due to various factors inversely affecting the production. Crop loss is happening because of the pest attacks and diseases [3-4]. In Kerala, Coconut Development Board started in the year 1981that comes under the Ministry of Agriculture and Farmers Welfare, Govt. of India with the aim of development in the coconut cultivation. The Board set up a Technology Development Centre at Aluva in Kerala provides





ambient support for the expansion of coconut industry in India. Coconut industry plays a significant role in foreign exchange. There are Coconut Producer Organizations formed in Kerala comprising Coconut Producer Societies (CPS), Coconut Producer Federations (CPF) and Coconut Producer Companies (CPC). CPS has 40 – 100 farmers as members dealing with procurement, processing and marketing phases of Coconut. Coconut Producer Company comprises of several Coconut Producer Federations and each CPF has many Coconut Producer Societies that directly interact with coconut farmers [5].

Various products that can be developed from coconut are available in the global market. Some includes [6-7]:

Desiccated Coconut: Desiccated coconut is used by confectionary and food industries in large quantity which is made up of grating the dehydrated coconut. Depending upon the fineness of the product, it is divided into a number of grades.

Coconut Milk: Coconut Milk deriving out of fresh grated coconut kernel is rich in protein can be used as coconut cream or diluted makes dishes such as puddings and sweets makes more healthy. Preserved forms of coconut milk are available in all countries. Fully defatted coconut milk is coconut cream, which can be used as an ingredient in house hold recipes and is available in global market. Coconut Development Board made new technologies to prepare coconut cream in a low cost mode.

Coconut Skimmed Milk: a soluble component is also prepared from coconut after removing cream using cream separator. Dried form can be kept as coconut milk powder. Nata-de-coco is a floating type dessert, a white creamy substance made from coconut. Other coconut products developed are coconut chips, squash, neera, jiggery, palm sugar, flower syrup and coconut toffee,

Coconut Oil: It is significant cooking oil apply in Kerala. Considering the peculiarity of low melting behavior and good digestibility, good scent oil is preferred most people. It has more medicinal values like shortening blood cholesterol and diminishes serum triglycerides. Among all vegetable oils, coconut oil has lowest unsaturated fatty acids. Milling copra is used for oil preparation as it contains highest percentage of oil than other oil seeds. The edible copra is a good dry fruit that can be used for confectionaries.

Tender Coconut Water: A health drink offered by Doctors that contain rich in potassium and minerals and is a thirst quenching health drink. 100 gram of tender coconut water has 17.4 caloric values. Tender coconut is a pre-maturing stage of fully matured nut.

Coconut Vinegar: A preservative agent made from fermented coconut water is coconut vinegar used in pickles, sauces and various food products. It contains minerals and vitamins and is natural than the synthetic vinegar.

Coconut Candy and Biscuits: By mixing coconut powder and maida in a convenient proportion make a ready to eat snack products biscuits

Coconut Development Board constituted in the state gives ambient technical advisory, training sessions and support for setting products based on coconut. Also quality testing criteria has





been incorporated to ensure the quality of the coconut product. CPCs installed high quality plants and machineries for the processing of neera and other coconut products like coconut honey and jiggery, making it an expert quality. New technologies such as pasteurization and chilling are used for preserving neera in CPCs.

In this paper, we are going to discuss various diseases found in coconut and related symptoms. The review pointed out different diseases found in various parts of different countries.

LITERATURE REVIEW

Most phytoplasma associated vector found in area of Jamaica is the pest named leafhopper and in Florida, insect named Haplaxius crudus.

The integrated disease management package is adopted in Jamaica to protect the coconut crop. The package consists of surveillance, in which the symptom should be reported as early as possible. If symptoms are found, eliminate those plants and replant with resistance effective one to keep the density. Make the plantation free of all types of weeds so that the plants could grow healthy and proper measure should be taken to control vectors [8-12]. Previous researches show that there are around 750 species types attacking coconut worldwide. Insect's pests causes severe damage that may incur losses in number and size of coconut [13].

Spreading is controlled by removing the infected trees and replanting with resistant variety trees. To control infection of phytoplasma in individual host plants, injecting antibiotic such as tetracycline is preferable but is not economically viable for small farmers. Climate disturbances impact the spreading of LYD and other phytoplasma associated diseases [14]. Phytoplasma 16SrIVA-F subgroups are found in 8 Mexican states.

In [15], the author implemented a Deep Learning based unsupervised learning algorithm to detect the pest attack on coconut palm. They used camera interfaced drone to collect images of coconut in the farm and then applied cropping technique for normalization and then created a database using NVIDA Tegra System on Chip. The database includes the images of coconut segments such as leaves, trunks and nuts. Smoothing followed by Image Enhancement technique used on the plant images. Healthy and unhealthy images and severity of the disease are classified along with the level of health based on the intensity and variation of green color. The GoPro camera mounted drone used for capturing images fly under the height of 400 feet which is in the line of sight of the operator [16].

Frequent Diseases Found in Coconut

1. LYD (Lethal Yellowing Disease)

Around more than 50 types of diseases have been effected by coconut worldwide. Lethal Yellowing and root wilt diseases are commonly found causes reduction in nut production in Kerala and thereby incur losses to farmers. Most of the diseases have the symptom of yellowing and necrosis of leaves [2, 17]. Many diseases attacked by coconut plant leads to low growth of tree and ultimately lead to stubby production. Major pest types attacking coconut are coconut caterpillar and lack beetle and minor pests are bagworm, coconut scale and leaf minors.





Previous research has been pointed out that diseases affect coconut production inversely and thereby a threat to coconut farmers [3, 8]. Lethal Yellowing is treated as the first phytoplasma related disease in coconut. The first visual symptom shown in coconut palms is the premature drop of fruit. No fruit set on inflorescences as most of the male flowers die during the second stage. With the development of necrosis in more than two inflorescences, leaves starts to become yellow. Yellowing of leaves begin from oldest one, and then spread to the middle one and finally the upper leaves. Yellow leaves then turns brown and finally die within a period of 3 - 6 months. Symptoms shown in one or two palms in the initial spread and randomly spread up to 100m in the secondary stage and jumped anywhere upto 100km in final stage [14-20, 53].

Vector transmission is the main cause of spreading LYD most probably through the movement of plant materials. Diagnosis of LY phytoplasmas was done using transmission electron microscopy in early days but it was more time consuming [21-22]. Dot blot analysis is a modified molecular technology that can detect more samples but the disadvantage is that the sensitivity and specificity were limited. Afterwards PCR technique has been introduced for phytoplasma detection, ensuring specificity and sensitivity [23-24].

2. Red Ring Diseases

Red ring diseases are commonly found in Trinidad and Tobago recent years and are known to be root disease. The vector named Bursaphelenchus cocophilus nematode is the main cause of Red Ring Disease. Symptom is that, a plumping appearance shown by shortening the leaves of the central package [21, 25]. In Ecuador, research has been conducted to find out the Bud Rot disease of coconut and they found VARI (Visible Atmospherically Resistant Index) as the best vegetation index for the early diagnosis of Bud Rot disease. They used high resolution multi spectral cameras for acquiring images of both healthy and diseased palm images for the analysis. Points of plants showing BR and RRD symptoms collected using GPS system and experts gone directly to the farm for analyzing the spatial behavior of the disease. Vegetation index shows the difference between healthy and diseased palms [26]. For diagnosing red ring disease, an identification procedure was applied, a Loop Mediated Isothermal Amplification (LAMP) to the causative agent *Bursaphelenchus cocophilus* [27]

Red ring disease have initial symptom within 14-21 days after infestation and shows the yellowing of oldest 2-3 leaflets and get maximum population of nematode in around 42 days and then petioles and roots also get infected and the leafs become bronze colored. Meanwhile nuts also fell down in a premature stage and red ring of 2-6cm has been formed. It has been found out that premature death of coconut palms at the age of 3-10 years occurred and older age palms, especially of 20years survive from red ring diseases. The end of monsoon and the starting of dry season affect inversely on palms and causes to the death of palms. It is found that Sugarcane weevils and American weevils are the nematodes spreading red ring disease [29-30]. Red ring nematode is found in some areas of South America, Central America and many Caribbean islands. Red ring nematodes block water pathways in leaves, roots and stem and new leaves become short. An integrated pest management system should be followed to control the insects, keeping hygene and minimization of pesticides [32].





3. Bud Rot Disease

Bud Rot is a most forbidding disease found in Ecuador, South Colombia and in South American Countries. Symptom is necrosis and disintegration of younger arrows with a yellow color and finally palms fell into sick and in unproductive state. The dreaded fungus named Phytophthora palmivora attacks results in bud rot disease [30]. In Bangladesh, study was conducted on different locations for an environmental friendly disease management method and the main diseases found in coconut were grey leaf spot or blight and bud rot disease. Weather factors such as temperature, rainfall and relative humidity and location of plant plays a major role for varying the disease [31]. Bud Rot effected palm shows the detachment of external tissues by leaving the secondary ribs. In the final stage of the disease, plant can't issue new arrows which indicate the total damage. Even though palms of all age are to be attacked, young ones are more susceptible. This disease is found in climate of humidity and low temperature such as monsoon season. Symptoms are black spots on the leaves followed by yellowing and finally spindle withers and drops down with a foul smell. Nuts are retained in their palms goes maturity.

From the previous studies, it is found that bud rot disease was commonly found in environment conditions of high humidity and low temperature. Heavy rains and cyclones increase the intensity of the disease in 2020-2022 year. Spreading of disease in the garden soil is carried out through communication and movement of human, birds, insects and snails, especially in rainy season [32]. Preventive measures should be taken to come down the disease such as proper spacing should be maintained and can apply copper oxy chloride 3g/lit of water [33].

4. Ganodarma Butt Rot

Ganoderma is a wood stinking fungus attacking old or weak palms, especially coconut palms throughout the world. It is also known as Basal Stem Rot (BSR) and is spread through soil. Species like *G. applanatum*, *G. boninense*, *G. lucidum*, *G. zonatum* are the causal agents of the disease. Even though Ganoderma species have important economic value, it has the dark side to spoil living trunks coconut palms. Symptoms shown are the diminishing of the palm from normal growth, becoming pale green color of the leaves and fungus kills the trunk and drops the entire palm while the disease progresses [34].

Lower leaves become yellow color and fine roots become decayed. Just near the ground level of the stem, bleeding patches appeared that extends upwards on the progress of the disease and parallel shedding of nuts occurs. Stem decay also takes place with a bad smell as fluid of reddish brown color comes from the stem and decay moves upwards. Meanwhile leaf production becomes reduced and also shortage of leaves turned out [35]. Young palms infected by Ganodarma disease die within 6-24 months after the first appearance of the symptom. Mature plants take some more time probably 2-3 more years to complete drop [36]. Two diagnostic methods available today for the diagnosis of Ganoderma are Pabs(Polyclonal antibodies) and PCR(Polymerase Chain Reaction) methods. Ganoderma survives in the soil for a long time. The transfer of proteins and nutrients, water irrigation causes the disease spreading as there are more fungus resides in a plant. It is also know by name Tanjore wilt, leaf blight found in Pollachi area of Tamilnadu. Survey conducted to assess in coconut growing districts





of Tamil Nadu [37].

Symptoms found on Tanjore wilt starts with withering, yellowing and drooping of the outer whorl of leaves and reddish brown liquid spread out from the stem and moves upward with a bad smell. Bracker forms at the base of the trunk in rainy seasons and finally palm dies [38-39].

5. Stem Bleeding

Thielaviopsis paradoxa (de Seyness) Von Hohnel is the cause of Stem bleeding disease. The symptom of Stem Bleeding starts with the emission of dark reddish liquid from the barks and turns black for a distance of not very many inches and speading upwards gradually as the disease progresses. When it damages stem tissue, leafs become yellow and drop one by one and nut falls prematurely and crown size become shrunk.

Back water soil and other sandy soils are found as more suitable for the spread of stem bleeding disease in coconut but also rarely found in all soil types. Also hard soil types of dry spells with high temperature circumstances are favors for the pathogen for increasing disease to an extend [40].

6. Leaf Rot

Water-soaked brownish lesions can be seen in the spear leaves at the earlier stage and then enlarges and get dry giving a fan shape is the symptom of the leaf rot affected palm. Unopened leaflets with pale yellow color are more susceptible to this disease. Wall less prokaryotes phytoplasma is the cause of leaf rot disease. [41-45].

7. Leaf Blight

It is a non-lethal, exhausting disease found in all parts of the country. In draught climatic conditions especially in summer season, the airborne disease is found out as leaf blight severely affecting the production of nuts in coconut plants. Initial symptom of the disease is the yellowish spots surrounded by grayish line appear on the older leaflets and burnt afterwards [46]. Weligama coconut leaf wilt disease spread by insect vector found in coconut of Sri Lanka

8. Root Wilt Disease (Kerala Wilt)

Symptoms found on root wilt disease are the flexibility of leaves curving inwards totally, found in palms having more than 30 months old. Yellowing of leaves followed by wilting, necrosis of leaflets and falling of nuts also occur. There is curtailment in amount of leaves as well as in size. Matured nuts seem to be smaller than normal and size of the crown get reduced. Roots also show symptoms like cracks and rotting [47-49].

It is a phytoplasma associated disease commonly found in South India causing economic loss and yield reduction. Phytoplasmas have wall-less cells and unculturable. Through insect vectors, pathogens transmit to other plants. It has different names in different areas like Weligama coconut leaf wilt in south region of SriLanka and Klimantan in Indonesia. The root wilt affected palm shows delaying in flowering. It was first observed in Kerala in the year 1882. It takes nearly 6-24 months to show the clear symptoms and take 3 years in rare cases [50].





9. Cadang-Cadang Disease

A lethal viroid named Coconut cadang-cadang viroid (CCCVd) is the source of lethal cadangcadang disease. Symptom shows very slowly. Just after the flowering, leaf shows yellow spots. Yellow spots spread everywhere in the initial stage, coconut production become very slow in the intermediate stage and in the final stage no more coconuts are produced at the end of 4-6 years after first symptom has been found out. Palms of less than 10 years are found to be less affected by cadang-cadang disease. CCCVd killed millions of palms in Philippines [51-52].

Viroid is a small single stranded RNA molecule and could not have protein coating with the property of self-replicating property. The already damaged palm by some insects is the place of viroids as they infiltrate cells of the damaged membrane. Viroids are resistant to high temperatures and it spread disease to healthy plants [53-54]. The spreading of virus may take plan through farm tools of poor sanitation

COMPARATIVE ANALYSIS

The listed diseases are commonly found in Asian countries like India, Indonesia, Malaysia, Philippines, Sri Lanka and in Thailand. Red ring disease is commonly found in Pakistan. Ganoderma, Leaf root and root wilt diseases are found in Bangladesh, Coconut in Myanmar is affected by Ganodarma disease. African countries such as Ghana, Ivory Coast, Nigeria, Tanzania, Mozambique and Kenya found almost all diseases except cadang-cadang. Lethel vellowing, red-ring, bud rot, leaf wilt and root wilt diseases are common in both central and south American places such as Costa Rica, Hodduras, Nicaragua, Panama and in Colombia, Guyana, Suriname, Venezuela and Brazil. Red ring disease was found in Ecuador of South America. Bud rot and Leaf rot are identified in Peru, a South American state. Carribbean islands such as Jamaica, St. Lucia, Trinidad and Tbago, Cuba, Haiti are famous for coconut plantations and various diseases such as lethal yellowing, bud rot, leaf rot and root wilt diseases are affecting their economic status of the country. Country, Papua New Guinea mainly found the ganodarma, cadang-cadang and leaf blight diseases. Stem bleeding is found in Fiii. Samoa. Solomon Islands and Vanuatu part of Oceania. Also Samoa, Indonesia and Malavsia of Oceania affected by the cadang-cadang disease. Asia, Africa, Central and South America, Caribbean and Oceania of the world countries cultivated coconut and the diseases affects inversely on the production of coconut. Table1 depicts common coconut diseases found in various part of the world along with their affected area of the coconut tree and the figure1 represents the plot on diseases vs countries.







Disease	Author	Location	Affected area
Lethal Yellowing Disease	[3,8,10- 20,22-25, 40]	 Asia: India, Indonesia, Malaysia, Philippines, Sri Lanka, Thailand Africa: Ghana, Ivory Coast, Nigeria, Tanzania, Mozambique, Kenya, Zanzibar Central America: Costa Rica, Honduras, Nicaragua, Panama South America: Colombia, Guyana, Suriname, Venezuela, Brazil Caribbean: Jamaica, Puerto Rico, St. Lucia, Trinidad and Tobago, Antigua and Barbuda, Bahamas, Barbados, Belize, Cuba, Dominical Republic, Grenada, Haiti, Guadeloup, Martinique, Monteserrat, St. Vincent and the Grenadines, Kitts and Nevis 	Leaf
Red ring	[21,28- 30,32]	Asia: India, Indonesia, Malaysia, Philippines, Sri Lanka, Thailand, Pakistan Africa: Ghana, Ivory Coast, Nigeria, Tanzania, Mozambique, Kenya, Zanzibar, Cameroon, Democratic Republic of Congo, Gabon, Togo Central America: Costa Rica, Honduras, Nicaragua, Panama, Guatemala, Mexico South America: Colombia, Guyana, Suriname, Venezuela, Brazil, Euador	trunk and leaves
Bud rot	[29, 34]	Asia: India, Indonesia, Malaysia, Philippines, Sri Lanka, Thailand Africa: Ghana, Ivory Coast, Nigeria, Tanzania, Mozambique Central America: Costa Rica, Honduras, Nicaragua, Panama, Guatemala, Mexico, Belize, El Salvador South America: Colombia, Guyana, Suriname, Venezuela, Brazil, Peru, Ecuador Caribbean: Jamaica, Puerto Rico, St. Lucia, Trinidad and Tobago, Antigua and Barbuda, Bahamas, Barbados, Belize, Cuba, Dominical Republic, Grenada, Haiti	buds
Ganodarma	[35-37]	Asia: India, Indonesia, Malaysia, Philippines, Sri Lanka, Thailand, Bangladesh, Myanmar Africa: Ghana, Ivory Coast, Nigeria, Tanzania, Cameron Oceania: Papua New Guinea	trunk
Stem Bleeding	[2,26,42]	Asia: India, Indonesia, Malaysia, Philippines, Sri Lanka, Thailand Africa: Ghana, Ivory Coast, Nigeria, Tanzania, Mozambique Oceania: Fiji, Samoa, Solomon Islands, Vanuatu	trunk
Leaf rot	[32,35,36]	 Asia: India, Indonesia, Malaysia, Philippines, Sri Lanka, Thailand, Bangladesh Africa: Ghana, Ivory Coast, Nigeria, Tanzania, Mozambique, Kenya Central America: Costa Rica, Honduras, Nicaragua, Panama South America: Colombia, Guyana, Suriname, Venezuela, Brazil, Peru, Ecuador 	leaves

Table 1: Coconut diseases found in different locations in various countries







		Caribbean: Jamaica, Puerto Rico, St. Lucia, Trinidad and Tobago, St. Vincent and the Grenadines	
Leaf blight	[26,32,50]	Asia: India, Indonesia, Malaysia, Philippines, Sri Lanka, Thailand Africa: Ghana, Ivory Coast, Tanzania, Mozambique, Kenya Oceania: Australia, Fiji, Papua New Guinea	leaves
Root wilt(kerala wilt)	[2,32,47,4 8]	Asia: India, Indonesia, Malaysia, Philippines, Thailand, Bangladesh Africa: Ghana, Nigeria, Tanzania, Mozambique Central America: Costa Rica, Honduras, Panama South America: Colombia, Guyana, Suriname, Venezuela, Peru Caribbean: Jamaica, Puerto Rico, St. Lucia, Trinidad and Tobago, Barbados, Dominican Republic, Grenada, Haiti	roots
Cadang- cadang	[30,51-54]	Asia: India, Indonesia, Malaysia, Philippines, Thailand Oceania: Papua New Guinea, Samoa, Indonesia, Malaysia	Growing point

Figure 1: Graphical representation on coconut diseases vs countries



CONCLUSION

As replanting of each palm is not economically viable, proper control measurements should be adopted to limit the diseases of coconut. This paper illustrates the common diseases found in coconut and it will help to take early adoption of control strategies to limit the economic loss of the country to some extent. Further research is required in the field to identify the early detection of disease and implement innovative disease management strategies for the sustainable crop production.





References

- 1) Megalingam RK, Manoharan SP, Garlapati K, Kota N, Mondreti YS, Gutha P, Tantravahi S. Analysis of Coconut Tree trunk for the Identification of Cracks and Holes Using Image Processing. In2022 IEEE 7th International conference for Convergence in Technology (I2CT) 2022 Apr 7, (pp. 1-6). IEEE.
- Reddy K V K, Rao G P, Singh A K, Baranwal V K, Singh B, "Detection of Phytoplasmas Associated with Wilt-Like Disease in Coconut Palms in India", PLoS ONE. 2021 Apr21;16(4) : e0250165. doi: 10.1371/journal.pone.0250165.
- Gurr GM, Johnson AC, Ash GJ, Wilson BA, Ero MM, Pilotti CA, Dewhurst CF, You MS. Coconut lethal yellowing diseases: a phytoplasma threat to palms of global economic and social significance. Frontiers in plant science. 2016 Oct 26;7:1521.
- 4) Kadethankar A, Sinha N, Burman A, Hegde V. Deep Learning Based Detection of Rhinoceros Beetle Infestation in Coconut Trees Using Drone Imagery. In Computer Vision and Image Processing: 5th International Conference, CVIP 2020, Prayagraj, India, December 4-6, 2020, Revised Selected Papers, Part I 5 2021 (pp. 463-474). Springer Singapore.
- Chaithanya KP, Melekoodappattu JG. An Exploration On Plant Disease Detection. In2022 Third International Conference on Intelligent Computing Instrumentation and Control Technologies (ICICICT) 2022 Aug 11 (pp. 911-916). IEEE.
- 6) S. Ekanayaka *et al.*, "IoT-Based Disease Diagnosis and Knowledge Dissemination System for Coconut Plants," *2022 4th International Conference on Advancements in Computing (ICAC)*, Colombo, Sri Lanka, 2022, pp. 126-131, doi: 10.1109/ICAC57685.2022.10025150.
- 7) K. H. R. Gunawardana, M. P. N. Deshan, M. G. S. P. Hemachandra, D. Ganegoda, N. M. Hettiarachchi and L. Weerasinghe, "Cocopal - A Deep Learning Based Intelligent System to Certify and Standardize the Quality of Coconut Based Products," 2022 4th International Conference on Advancements in Computing (ICAC), Colombo, Sri Lanka, 2022, pp. 470-475, doi: 10.1109/ICAC57685.2022.10025099.
- Aidoo OF, Cunze S, Guimapi RA, Arhin L, Ablormeti FK, Tettey E, Dampare F, Afram Y, Bonsu O, Obeng J, Lutuf H. Lethal yellowing disease: insights from predicting potential distribution under different climate change scenarios. Journal of Plant Diseases and Protection. 2021 Oct;128(5):1313-25.
- 9) Lebsky V, Hernandez J, Barraza A, Oropeza C, Narvaez M, Carrillo A, Castellanos T, Poghosyan A. Ultrastructural analysis of spiroplasmas detected in palm species infected with the lethal yellowing phytoplasma from Yucatan and Baja California Sur, Mexico. Phytopathogenic Mollicutes. 2019(1)
- 10) Córdova I, Oropeza C, Puch-Hau C, Harrison N, Collí-Rodríguez A, Narvaez M, Nic-Matos G, Reyes C, Sáenz L. A real-time PCR assay for detection of coconut lethal yellowing phytoplasmas of group 16SrIV subgroups A, D and E found in the Americas. Journal of Plant Pathology. 2014 Jul 5;96(2):343-52.
- 11) Contaldo N, D'Amico G, Paltrinieri S, Diallo HA, Bertaccini A, Rosete YA. Molecular and biological characterization of phytoplasmas from coconut palms affected by the lethal yellowing disease in Africa. Microbiological research. 2019 Jun 1;223:51-7.
- 12) Arocha-Rosete Y, Konan Konan JL, Diallo AH, Allou K, Scott JA. Identification and molecular characterization of the phytoplasma associated with a lethal yellowing-type disease of coconut in Côte d'Ivoire. Canadian Journal of Plant Pathology. 2014 Apr 3;36(2):141-50.
- 13) Myrie WA, Harrison NA, Douglas L, Helmick E, Gore-Francis J, Oropeza C, McLaughlin WA. First report of lethal yellowing disease associated with subgroup 16SrIV-A phytoplasmas in Antigua, West Indies. New Disease Reports. 2014 Apr 14
- 14) Tsai JH. Lethal yellowing of palms. InPlant Diseases of Viral, Viroid, Mycoplasma and Uncertain Etiology 2019 Sep 11 (pp. 83-94). CRC Press.





- 15) Alouw JC, Wulandari S. Present status and outlook of coconut development in Indonesia. InIOP Conference Series: Earth and Environmental Science 2020 (Vol. 418, No. 1, p. 012035). IOP Publishing.
- 16) Oropeza-Salín C, Sáenz L, Narvaez M, Nic-Matos G, Córdova I, Myrie W, Ortíz CF, Ramos E. Dealing with lethal yellowing and related diseases in coconut. Coconut Biotechnology: Towards the Sustainability of the 'Tree of Life'. 2020:169-97.
- 17) Amponsah-Mensah K, Teng P S, Wei W, et al, "Lethal yellowing disease of coconut: A review of past and current research efforts". Front Plant Sci, 2019;10:1508. Available from: https://doi.org/10.3389/fpls.2019.01508
- 18) Mwakasegele GK, Chang'a EP, Mneney EE, et al, "A bioinformatics approach for predicting effector proteins in phytoplasma causing lethal yellowing disease in coconut". BMC Bioinformatics, 2019;20(Suppl 25):670, https://doi.org/10.1186/s12859-019-3215-6
- 19) Klassen S P, Menzel C M, Chinchilla C M, et al, "Transmission and impacts of lethal yellowing-like diseases of coconut, occurring in the Western Hemisphere: A review", PLoS One. 2020;15(1):e0227642, https://doi.org/10.1371/journal.pone.0227642
- 20) Kusi F, Adu-Acheampong R, Agyei-Dwarko D, et al. Distribution of lethal yellowing disease of coconut palm in Ghana: An overview of phytoplasma strains. Afr J Agric Res. 2019;14(15):725-735
- 21) Li X, Wang J, Li H, et al, "Red ring disease detection in coconut trees using unmanned aerial vehicles and deep learning", IEEE Access, 2021;9:19968-19977, https://doi.org/10.1109/access.2021.3050548
- 22) Marraiki N, Al-Zarouni M, Soliman M, et al, "Detection and characterization of phytoplasma associated with lethal yellowing disease of coconut in the United Arab Emirates", Int J Bioinform Res Appl, 2019;15(3):288-300. Available from: https://doi.org/10.1504/ijbra.2019.099175
- 23) Sato S, Pathak P D, Meshram M, et al, "Detection of lethal yellowing phytoplasma in coconut leaves by loop-mediated isothermal amplification (LAMP) and SYBR Green I-based real-time PCR", PLoS One, 2019;14(6):e0218132. Available from: https://doi.org/10.1371/journal.pone.0218132
- 24) Gerber K, Du Plessis H, Kvarnheden A, et al, "Lethal yellowing disease: a rapid review of current knowledge and research needs", Front Plant Sci [Internet]. 2017 Nov 9;8:1952. Available from: https://doi.org/10.3389/fpls.2017.01952
- 25) Wang X, Wang L, Jiang L, et al, "Early detection of lethal yellowing disease in coconut using machine learning algorithms". IEEE Access, 2021;9:54903-54910. Available from: https://doi.org/10.1109/access.2021.3078194
- 26) Chandy A. Pest infestation identification in coconut trees using deep learning. Journal of Artificial Intelligence. 2019 Sep;1(01):10-8.
- 27) Alshawwa IA, Elsharif AA, Abu-Naser SS. An Expert System for Coconut Diseases Diagnosis.
- 28) Martyn EE. Red ring disease of coconut in Trinidad and Tobago, Tropical Agriculture. 2021 Sep 29.
- 29) Viera-Torres M, Sinde-González I, Gil-Docampo M, Bravo-Yandún V, Toulkeridis T. Generating the baseline in the early detection of bud rot and red ring disease in oil palms by geospatial technologies. Remote Sensing. 2020 Oct 3;12(19):3229.
- 30) Arif MI, Rafique M, Siddique MT, et al. Characterization of phytoplasma associated with red ring disease in coconut palms in Pakistan. Phytopathol Mediterr. 2018;57(3):463-471
- 31) Geistlinger J, Rath T, Baumert A, et al, "The complete nucleotide sequence of coconut cadang-cadang viroid, a member of a novel group of viroids". Front Microbiol, 2017 Feb 24;8:262, https://doi.org/10.3389/fmicb.2017.00262]





- 32) Ide T, Kanzaki N, Giraldo PP, Giblin-Davis RM. Loop-mediated isothermal amplification (LAMP) for detection of the red ring nematode, Bursaphelenchus cocophilus. Nematology. 2017 Jan 1;19(5):559-65.
- 33) Khan MA, Hossain I. Leaf spot disease of coconut seedling and its eco-friendly management. Journal of the Bangladesh Agricultural University. 2013;11(452-2016-35595):199-208.
- Vinjusha N, Arun Kumar TK. Revision of Ganoderma species associated with stem rot of coconut palm. Mycologia. 2022 Jan 2;114(1):157-74.
- 35) Nair GA, Abirami K, Varghese S, et al, "In-silico analysis of Serine/Threonine protein kinase genes in Phytophthora palmivora, the causal agent of bud rot disease in coconut", Int J Bioinform Res Appl, 2018;14(2):166-74. https://doi.org/10.1504/ijbra.2018.090202
- 36) Ahmadi P, Muharam FM, Ahmad K, Mansor S, Abu Seman I. Early detection of Ganoderma basal stem rot of oil palms using artificial neural network spectral analysis. Plant disease. 2017 Jun 12;101(6):1009-16.
- 37) Bisoyi SK, Das S, DebasisPattnaik A, Srivastava S. Ganoderma lucidum: As Calamitous and Salutary Mushroom.
- 38) Zulperi D, Zakaria L, Ismail N, et al, "Transcriptomic response of oil palm (Elaeis guineensis Jacq.) to the infection of Ganoderma boninense", BMC Bioinformatics, 2019 May 10;20(Suppl 7):184, https://doi.org/10.1186/s12859-019-2694-4
- 39) Vinayaka H, Prathibha VH. Integrated disease management in coconut. Indian Coconut Journal. 2013;56(3):16-21.
- 40) Elliott ML, Uchida JY. Diseases and disorders of ornamental palms. Phytopathology News. 2022 Jan.
- 41) Perera L, Kulasekera L, Gunasinghe U B, et al., "Pathway analysis and biological interpretation of long noncoding RNAs identified in a genetic screen for coconut lethal yellowing disease", BMC Bioinformatics, 2020 Jun 3;21(1):233, https://doi.org/10.1186/s12859-020-03564-w
- 42) Ramjegathesh R, Karthikeyan G, Rajendran L, Johnson I, Raguchander T, Samiyappan R. Root (wilt) disease of coconut palms in South Asia–an overview. Archives of Phytopathology and Plant Protection. 2012 Dec 1;45(20):2485-93.
- 43) Karuppiah P, Prabukumar G, Raguchander T, et al., "Isolation and identification of pathogenic fungi causing stem bleeding disease of coconut in South India". Int J Bioinform Res Appl, 2016;12(3):287-96, https://doi.org/10.1504/ijbra.2016.077710]
- 44) Ramli MN, Abdullah SNA, Md Yusoff F, et al., "Pathogenicity of Phytophthora palmivora and P. capsici on coconut (Cocos nucifera L.) roots and leaves", Front Plant Sci, 2017 Mar 3;8:289, https://doi.org/10.3389/fpls.2017.00289
- 45) Ravanbakhsh S, Liu Z, Akoglu A, et al, "DeepPheno: Predicting single-gene knockout phenotypes using an attention-based deep learning approach on imaging data", BMC Bioinformatics [Internet]. 2019 Nov 14;20(1):599, https://doi.org/10.1186/s12859-019-3227-1
- 46) Elanchezhian R, Justin C C, Gopalasundaram P, "Molecular characterization and phylogenetic analysis of Colletotrichum gloeosporioides, the causal agent of leaf rot disease in coconut", Int J Bioinform Res Appl, 2015;11(5):496-507, https://doi.org/10.1504/ijbra.2015.072832
- 47) Hassan MM, Jahan S N, Molla AH, et al., "Identification and characterization of fungal pathogens causing leaf rot disease of coconut (Cocos nucifera L.) in Bangladesh", Int J Bioinform Res Appl, 2021;17(3):254-69, https://doi.org/10.1504/ijbra.2021.114045]
- 48) Manimekalai R, Soumya VP, Nair S, Thomas GV, Baranwal VK. Molecular characterization identifies 16SrXI-B group phytoplasma ('Candidatus Phytoplasma oryzae'-related strain) associated with root wilt







disease of coconut in India. Scientia Horticulturae. 2014 Jan 22;165:288-94.

- 49) Nair S, Manimekalai R, Ganga Raj P, Hegde V. Loop mediated isothermal amplification (LAMP) assay for detection of coconut root wilt disease and arecanut yellow leaf disease phytoplasma. World Journal of Microbiology and Biotechnology. 2016 Jul;32:1-7.
- 50) Mwirichia J M, Chen C, Ngumbi P M, et al., "Metagenomic analysis of the microbial community associated with root (wilt) disease of coconut palm in Tanzania", PLoS ONE [Internet]. 2018 Apr 12;13(4):e0191914, https://doi.org/10.1371/journal.pone.0191914
- 51) Ashokkumar P, Ushamalini C, Ramjegathesh R. Variations in Morphological and Molecular Characterization of Lasiodiplodia theobromae (Pat.) Griffon and Maubl Associated with Coconut Leaf Blight. Madras Agricultural Journal. 2018 Dec 20;105(march (1-3)):1.
- 52) Hanold D, Randles JW. Coconut cadang-cadang disease and its viroid agent. Plant Disease. 1991 Apr 1;75(4):330-5.
- 53) Acedo AL, Oña J R, Villegas V P, et al., "Detection of cadang-cadang viroid in coconut (Cocos nucifera L.) using reverse transcription loop-mediated isothermal amplification", IEEE Access. 2020;8:184624-184633. Available from: https://doi.org/10.1109/access.2020.3031478
- 54) Shylaja, & Hemalatha N. (2023). A Review Of Computational Biology Approaches In Agriculture. *Redshine Archive*, *1*. https://doi.org/10.25215/8119070682.01
- 55) Madihah AZ, Maizatul-Suriza M, Idris AS. Reverse transcription loop-mediated isothermal amplification (RT-LAMP) for detection of Coconut Cadang-cadang Viroid (CCCVd) variants in oil palm. Journal of Oil Palm Research. 2020 Sep 1;32(3):453-63.
- 56) Umadhay K, Santos V, Silaran M, et al., "Host and non-host plant species as sources of inoculum for cadangcadang viroid in coconut", Front Microbiol, 2017 Jul 11;8:1289. https://doi.org/10.3389/fmicb.2017.01289.

