

## EFFICIENCY OF *PRIESTIA ARYABHATAI* COMBINATION WITH DIFFERENT NITROGEN FERTILIZER ON GROWTH, YIELD ATTRIBUTE, YIELD AND QUALITY OF BABY MAIZE (*ZEA MAYS L.*)

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### Abstract

The efficiency of five various nitrogen ratios (0, 325, 224.8, 162.5 and 81.3 kg N/ha) and *Priestia aryabhatai* on soil chemical properties, growth, development and productivity of baby corn was carried out outside net house in agricultural research center of An Giang university from Jun to August of 2023. The experimental design was performed by using randomized complete block design with five treatments and four repeats. Research results showed that various nitrogen ratios remarkably increased on soil chemical properties (Available P, total N, CEC and OM), biomass and ear number per plant. Furthermore, nitrogen rates and *Priestia aryabhatai* were significantly affected plant height, leaf number and total chlorophyll per plant. Nonetheless, five N<sub>2</sub> ratios applied, the optimal N level between 162.5 and 243.8 kg N /ha combined with *Priestia aryabhatai* inoculation potentially raised the edible cob yield, indicating that N treatment had the ability to produce higher edible cob yield as well as higher quality contents of baby corn edible cob. However, Interaction between nitrogen rates and *Priestia aryabhatai* was not affected on yield composition, edible cob yield and edible cob quality of baby corn.

**Keywords:** *Priestia Aryabhatai*, Nitrogen Fertilizer, Baby Corn, Edible Cob, Quality, Fresh Yield

### INTRODUCTION

The baby maize has been planting as good choice for increasing the profit of tillers. Baby maize, which is the sweet flavor and harvest from 65 to 75 days after sowing, is a crop kind and provides green ears. Baby maize is an importance role as vegetable and salad in meals around the world. The baby maize that is a short duration, helps tillers to take up many crop yearly as intercrop other plants and helps the tillers to get more profit per unit area per unit time by increasing cropping output (Rani et al., 2015). Baby maize for vegetable aim is successfully planted in Asian countries like Vietnam, Thailand, Taiwan, etc. It has been brought into a good income because of its potential product for export and a good foodstuff (Gondaliya et al., 2022). The N<sub>2</sub> element plays an important role in different physiological function of baby corn. It raises the leaf width effectively, reducing senescence and essential element for creation of baby corn ear and kernel. Furthermore, N<sub>2</sub> element significantly improves maize yield (Torbert et al., 2011), promotes kernel's function by raising number and weight of the grain, affecting the final size of kernel (Hopf et al., 1992; John & Schmitt, 2007). The positive effects of N<sub>2</sub> on essential agronomic and yield components of baby maize has been observed by several prior studies (McCullough et al., 1994; Evans, 2008). Among the different generous nutrition required for various plants, The N<sub>2</sub> element, which has an important role and particularly, has been discovered for maize by many various researches (Subramanian et al., 2006; Carpici et

al., 2010). The N<sub>2</sub> demand of baby corn is to depend on conditions of soil kinds, weather and crop rotation system (Blackmer, et al., 2009; Bundy, et al., 2011).

The growth and productivity of plants mainly relies on the interactions with rhizosphere microorganisms, which exist the crop soil. These relationships are very complicated and critical for meeting the bio-difference per the soil type (Lau and Lennon, 2011; Bever et al., 2013; Shao et al., 2018). Microorganisms are the most abundant and diverse entity in soil and can be directly involved in ecological processes and nutrient cycling. The rhizosphere microorganism role of agricultural soil has been significantly studied via isolation and identify of cultural microbes and culture-independent techniques (Liu et al., 2019; Roy et al., 2020). According to studied results of Nguyen Van Chuong et al., (2023) presented that co-application of vermicompost with three rhizosphere N<sub>2</sub> fixing bacterial strains raised the growth, yield attribute and yield of peanut to compare with inorganic fertilizers alone. further, addition of vermicompost and with *E. asburiae* inoculation, which obtained the maximum yield attributes and yield of peanut when planting on the poor nutrition soil, produced better increase in number and weight of peanut nodules compared with chemical fertilizer application alone. The research discovered the effects of rhizosphere microbial and soil nutrients to raise yield of various crops under sustainable agriculture systems. Roots contain a huge biomass and the important part of the crops to take water and nutrients for the plant growth. Rhizosphere and the narrow soil nearly contact the root is the plant root-soil interface and is the hot spot for interactions between plants and rhizosphere bacteria (Nguyen Van Chuong, 2023; Korenblum et al., 2020). Rhizosphere N<sub>2</sub> fixing bacteria and the crop could form symbiotic interactions where the root microorganism use root exudates and secretes compositions positive plant growth (Berendsen et al., 2012). Rhizosphere N<sub>2</sub> fixing bacteria can synthesize antibiotics for prevention of soil-borne pathogens and protect the plant health (Mendes et al., 2013; Lazcano et al., 2021). Other interactions between rhizosphere bacteria and plant consists of N<sub>2</sub> fixing bacteria and contributing essential elements (e.g., nitrogen) for growth and yield of plant (Moreau et al., 2019). Rhizosphere N<sub>2</sub>-fixing microbiome could also bring root exudation of metabolites and mediate root-root signaling promoting soil conditioning (Korenblum et al., 2020). Rhizosphere microorganisms could have ability of hormone production, which helps to promote growth and nutrient uptake of plant, inhibit and kill the growth of pathogenic bacteria and create crop resistance to biological or no biological stresses (Ahmed and Hasnain, 2014).

One of the most researched genera is *Bacillus* strains among the benefit rhizobacteria for plant growth and and shows huge potential in raising the growth and yield of crops(Chen et al., 2007; Tahir et al., 2017; Backer et al., 2018). *Priestia aryabhatai* (*Bacillus aryabhatai*) are widely existed in nature but were only discovered in 2009 (Shivaji et al., 2009). More strains of *B. aryabhatai* have been isolated from different plant roots, and appreciation of these rhizosphere bacterial strains have significantly related promising attributes of this *Bacillus* strains for the impulsion of plant growth and crop yield, further having drawn thriving of fascination from scientists (Bhattacharyya et al., 2017; Park et al., 2017; Ghosh et al., 2018). According to prior study of Mehmood et al. (2021) discovered that *B. aryabhatai* fostered wheat growth and decreased the impacts of stress on wheat due to salt concentration. Nonetheless, the rhizobia strain mechanism fostering plant growth and

development remains to have been studying. With new technologies, genome sequencing usage for identifying the isolated microbial was widely used to find out the new and potential species of the microbes in foster the plant performance (Chu et al., 2020; Chen et al., 2022). In a recent discovery, an excellently endogenous bacterium species, *B. aryabhatai*, was isolated from baby corn roots, and its effect was also found out in the recent study (Deng et al., 2022). Ability of *B. aryabhatai* has significantly fostered the plant growth and yield, such as N<sub>2</sub> fixing and phosphorus solution and IAA producibility. Objective of this study is to find out the best N<sub>2</sub> fertilizer rate combined with *B. aryabhatai* for the best baby corn growth and yield.

## MATERIALS AND METHODS

### *Priestia aryabhatai* resource

*Priestia aryabhatai* was **isolated** on aseptically nutrient agar (Nfb) media **and identified by** 16S rRNA sequencing technology **from the baby maize roots**. The baby corn root samples were collected from local farmers' farms in Cho Moi, An Giang, and Vietnam. *Priestia aryabhatai* was isolated in the center laboratory of An Giang university, and was identified through sequencing technology of 16S rRNA and phylogenetic position, were used by blasting the 16S rRNA sequence on NCBI. The similar rates of the 16S rRNA sequence of our target bacterium was 100% of the 16S rRNA sequence of our target bacterium (Chuong et al., 2023). *Priestia aryabhatai* was increased population on dilute nutrient Nfb media to be 10<sup>8</sup> CFU/ml, and then inoculated with baby corn seeds about 24 hours in dark before sowing.

### Design and location of experiment

A field experiment was carried out from Jun to august of 2023 outside the net house of agricultural research center in AG university. The field experiment consisted of five treatments (Table 1) with four N<sub>2</sub> fertilizer levels (0, 325, 224.8, 162.25 and 81.3 kg/ha) and four treatments (2, 3, 4 and 5) of *Priestia aryabhatai* inoculation (except control treatment) with four replications. The variety "HM-4" of baby corn was selected as a test crop. And four replications. The distance of each hole was 30 cm x 20 cm (166,666 plants/ha) and planted 03 seeds per hole. The variety Baby Corn 271 of Vino Joint Stock Company was planted during the experiment. Breed characteristics: milky white fruit, growing time from 50-55 days, healthy growth, high yield, good water tolerance. The whole area of study was 200 m<sup>2</sup> (1 m in width x 10 m in length x 04 replications x 05 treatments). Baby corn seeds preparation and bacterial population increase: baby corn seeds were incubated under dark conditions for germination one 24 hours before sowing. Then, baby corn seeds were well inoculated with a 10 mL bacterial (10<sup>8</sup> CFU/ mL) before sowing and two seeds per hole. Soil samples were taken 0-20 cm in the soil depth to determine the soil properties before experiment. Soil samples determined the physical – chemical properties by methods of Carter & Gregoric, (2007).

**Table 1: Nitrogen rates and *Priestia aryabhattai* inoculation of the experiment**

Treatments	<i>Priestia aryabhattai</i> (10 <sup>8</sup> CFU/ml)	Inorganic fertilizers (Kg/ha)		
		Nitrogen (N)	Phosphorous (P)	Potassium (K)
1 (control: 0% of N <sub>2</sub> )	Uninoculated	0.00	355	75
2 (100% of N <sub>2</sub> )	Inoculated	325		
3 (75% of N <sub>2</sub> )	Inoculated	224.8		
4 (50% of N <sub>2</sub> )	Inoculated	162.5		
5 (25% of N <sub>2</sub> )	Inoculated	81.3		

The stage of withdrawing the corn tassel plays an importantly role for growing baby corn ears, especially bringing high yield, focusing on nutrition for faster growing baby corn, shorter growth time, and increasing edible cob weight. Usually harvest is withdrawn the baby corn tassel from 45 to 50 days after sowing. Agronomy, yield attributes and edible cob yield, which were observed by during growth and development time of baby corn, counted such as height, number of branch, total chlorophyll, cob diameter and length of cobs, biomass, fresh weight of pods, corn silks, husks and cob per plant. The fresh yield of baby corn parts was counted by t/ha. Physical-Chemical analysis of the soil (top 20 cm) had a neutral pH (6.7), 0.80% SOM, 0.065 % total nitrogen, 58.1 mg/100g available phosphorus and no exchangeable potassium. Nitrogen as per the treatments was applied in three splits. The whole dose of P was applied one day before sowing, different nitrogen levels along with 80 kg K/ha was divided to apply at four stage from sowing to harvest. In general, experimental soil had very low nutrition and silt sand (sand: 80.0%, silt: 18.7%, clay: 1.3%).

### Statistical analysis

The recorded data of statistical analysis per character has been using the standard analysis of variance in split plot design ( $P < 0.05$ ) with the help of statistical software stat graphics software version XV.

## RESULTS AND DISCUSSION

### Effects of four N<sub>2</sub> ratios and *Priestia aryabhattai* inoculation on the Soil attributes at harvest

#### Soil pH

The results in Table 2 showed that soil pH ranged from 6.47 to 7.22 at all treatment and insignificant differences at level 5% at harvest. According to study of Ren et al., (2015) showed that there was not any effect of rhizosphere N<sub>2</sub> fixing bacteria and N<sub>2</sub> fertilizer on soil pH, was no their interaction on the soil pH.

#### Total nitrogen

The results in Table 2 showed that the total N<sub>2</sub> concentration of the soil at harvest ranged from 0.04 to 0.09 (%) in all treatments and was statistically significant differences at level 1%. The highest total N<sub>2</sub> content was 0.09 % at co-application of 162.5 kg N/ha + *Priestia aryabhattai* inoculation, and lowest total N<sub>2</sub> value (0.04 %) obtained at two treatments of 162.5 kg N/ha +

*Priestia aryabhatai* inoculation and 325 kg N/ha + *Priestia aryabhatai* inoculation (without nitrogen application). Entophytic bacteria play an important role in N<sub>2</sub> fixing processes of taking N<sub>2</sub> from the air into soils (NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub><sup>-</sup>) (Peng et al., 2022). Maize roots plant in a low N<sub>2</sub> condition that can stimulate N<sub>2</sub> fixing from the air and promote the N<sub>2</sub> fixation efficiency of maize by entophytic bacteria (Fan et al., 2019). Furthermore, levels of N<sub>2</sub> application affected a huge impact on rhizosphere N<sub>2</sub> fixing microorganisms and their community development. Some researchers have found that soil total nitrogen has a significant impact on bacterial community population (Zhang et al., 2020).

**Table 2: Chemical properties of soil at harvest**

Treatment	Chemical attributes of soil at harvest				
	pH H <sub>2</sub> O	Total N (%)	Available P (mg/100g)	CEC (Cmol <sup>+</sup> /kg)	OM (%)
0.00 kg N/ha + None <i>Priestia aryabhatai</i> inoculation	6.69	0.060 <sup>b</sup>	57.3 <sup>a</sup>	4.22 <sup>bc</sup>	0.880 <sup>b</sup>
325 kg N/ha + <i>Priestia aryabhatai</i> inoculation	7.22	0.040 <sup>c</sup>	39.2 <sup>b</sup>	4.99 <sup>b</sup>	0.950 <sup>b</sup>
224.8 kg N/ha + <i>Priestia aryabhatai</i> inoculation	6.68	0.040 <sup>c</sup>	51.0 <sup>a</sup>	4.43 <sup>bc</sup>	1.19 <sup>a</sup>
162.5 kg N/ha + <i>Priestia aryabhatai</i> inoculation	6.89	0.090 <sup>a</sup>	55.3 <sup>a</sup>	4.21 <sup>c</sup>	1.29 <sup>a</sup>
81.3 kg N/ha + <i>Priestia aryabhatai</i> inoculation	6.47	0.070 <sup>b</sup>	51.0 <sup>a</sup>	6.22 <sup>a</sup>	0.930 <sup>b</sup>
F	ns	**	*	**	**
CV (%)	8.85	14.4	17.3	18.8	18.1

The different letters in the same column indicate significant differences at 1% (\*\*), and insignificant difference at 5% (ns); DAS: days after sowing; CV: coefficient of variation.

#### Available phosphorous, CEC and Organic matter (OM)

Nitrogen rates (p<0.05), CEC (p<0.01) and their interaction (p<0.01) had remarkably affected on available P and CEC of crop soil (Table 2). The values of available P and CEC ranged from 39.2 to 57.3 mg/100g and 4.21 to 6.22 (Cmol<sup>+</sup>/kg), respectively. Available P, which was recorded in the co-application of 325 kgN/ ha and *Priestia aryabhatai* inoculation, had lower content than other treatments, while the minimum CEC value of 162.5 kg N/ha + *Priestia aryabhatai* inoculation. Relationship between N<sub>2</sub> level and N<sub>2</sub> fixing bacteria had less impact on available P and CEC of soil (Xia et al., 2020; Ran et al., 2021). The results in Table 2 showed that the soil OM at harvest ranged from 0.88 to 1.29% at all treatments and their interaction (p<0.01). Soil OM of 162.5 kg N/ha + *Priestia aryabhatai* inoculation was the highest OM content in 350 kg N/ha and lowest OM value in control. Leu, (2005) and Monaco et al. (2008), proved that interaction between entophytic N<sub>2</sub>fixiation bacteria and plant released over fifty percent of the total CO<sub>2</sub> released from rhizosphere soil. Soil organic matter are the increase or decrease in order to rhizosphere bacteria (Bade and Cheng, 2007), and is the most important factor of rhizosphere bacteria and root correlation (Bengtson et al., 2012).

**Effects of N<sub>2</sub> fertilizer rates and *Priestia aryabhatai* on agronomical composition**

**Table 3: agronomical attributes of baby corn**

Treatment	Plant height (cm)		Leaf number (leaf/plant)		Total chlorophyll (µg /mL)	
	15 DAS	30 DAS	15 DAS	30 DAS	15 DAS	30 DAS
1	33.5 <sup>b</sup>	95.8	6.50 <sup>c</sup>	12.5 <sup>ab</sup>	34.5 <sup>c</sup>	34.9 <sup>c</sup>
2	42.8 <sup>a</sup>	97.7	7.75 <sup>ab</sup>	12.8 <sup>a</sup>	43.3 <sup>a</sup>	39.8 <sup>b</sup>
3	40.4 <sup>a</sup>	96.0	7.00 <sup>bc</sup>	13.3 <sup>a</sup>	37.4 <sup>bc</sup>	44.0 <sup>a</sup>
4	38.8 <sup>ab</sup>	102	7.50 <sup>ab</sup>	13.3 <sup>a</sup>	39.4 <sup>ab</sup>	39.8 <sup>b</sup>
5	41.6 <sup>a</sup>	90.9	8.00 <sup>a</sup>	11.8 <sup>b</sup>	36.9 <sup>bc</sup>	39.7 <sup>b</sup>
F	*	ns	*	*	**	**
CV (%)	12.4	6.70	10.1	6.31	10.4	8.84

The different letters in the same column indicate significant differences at 5% (\*), at 1% (\*\*), and insignificant difference at 5% (ns); DAS: days after sowing; CV: coefficient of variation.

Studied data in Table 3 also presented that N<sub>2</sub> fertilizer ratios of 325, 224.8, 162.5 and 81.3 kg N/ha + *Priestia aryabhatai* inoculation were higher plant height as compared to control at 15 DAS, and their interaction at P < 0.05. Nonetheless, plant heights were not the interaction at 30 DAS. The leaf number and total chlorophyll were significant differences at level 5% (p < 0.05) and 1% (p < 0.01) at 15 and 30 DAS. Application of four N<sub>2</sub> fertilizer ratios (325, 224.8, 162.5 and 81.3 kg N/ha) combined with *Priestia aryabhatai* inoculation had higher leaf number and total chlorophyll index as compared to control treatment at 15 and 30 DAS. However, the highest leaf number (13.3 leaf/plant) and total chlorophyll (44.0 µg /mL) obtained in treatment 3 of 224.8 kg N/ha + *Priestia aryabhatai* inoculation at 30 DAS, whereas the lowest leaf number and total chlorophyll of control (treatment 1). The reduction in N<sub>2</sub> application with without *Priestia aryabhatai* inoculation could cause the intensified nutrient competition between plants and bacteria (Moosavi et al., 2012). Usage of right and sufficient nitrogen for increasing cod yield, profit's tillers and N<sub>2</sub> efficiency usage, while it minimizes the potential for loss of N<sub>2</sub>, thus decreasing environmental contamination. Nitrogen application has significant impact on growth yield of baby corn (Thavaprakash and Velayudham, 2009). Furthermore, suitable application of N<sub>2</sub> fertilizer could promote N<sub>2</sub> fixing process of rhizosphere bacteria due to N<sub>2</sub> deficiently of crop soils (Kar et al., 2006)

Effects of N<sub>2</sub> fertilizer rates and *Priestia aryabhatai* on yield attributes, yield and quality of baby corn Results in Table 4 showed that application of lesser N<sub>2</sub> fertilizer level (0.00 kg N/ ha) was significantly lower number of ears and biomass by 2.58 ear/plant and 26.8 t/ha, respectively, over higher N<sub>2</sub> levels (243.8 and 325 kg N/ha). The results of Table 4 also showed that, among all inoculated treatments of *Priestia aryabhatai* had number of ears (4.25 ears/plant) and biomass (38.8 t/ha) were higher than those of all uninoculated treatments at number of ears (3.74 ears/plant) and biomass (31.8 t/ha). Nitrogen rates (p < 0.01), and *Priestia aryabhatai* (p < 0.01), nonetheless, their interaction between the N<sub>2</sub> ratios and *Priestia aryabhatai* was insignificant impact on increasing number of ears and biomass (Table 4). Agronomical and yield attributes was significantly impacted by soil nutrient and types, planting technology, bacterial population, and environmental condition and weather, etc. (Zhai et al., 2019).

**Table 4: Yield Attributes and Yield of Baby Corn**

Factor	Number of ears (ears/plant)	Fresh yield (t/ha)				
		Ear	Silk	Husk	Edible cob	Biomass
<b>Nitrogen ratios (kg/ha) (A)</b>						
0.00	2.58 <sup>d</sup>	6.02 <sup>c</sup>	0.831 <sup>c</sup>	3.80 <sup>c</sup>	1.39 <sup>b</sup>	26.1 <sup>c</sup>
325	5.06 <sup>a</sup>	14.5 <sup>a</sup>	2.18 <sup>a</sup>	10.1 <sup>a</sup>	2.15 <sup>ab</sup>	35.1 <sup>b</sup>
243.8	4.43 <sup>b</sup>	15.0 <sup>a</sup>	1.85 <sup>b</sup>	10.4 <sup>a</sup>	2.75 <sup>a</sup>	42.8 <sup>a</sup>
162.5	4.38 <sup>b</sup>	10.5 <sup>b</sup>	0.950 <sup>c</sup>	6.53 <sup>b</sup>	3.00 <sup>a</sup>	35.1 <sup>b</sup>
81.3	3.50 <sup>c</sup>	9.60 <sup>b</sup>	0.950 <sup>c</sup>	6.73 <sup>b</sup>	1.93 <sup>ab</sup>	37.5 <sup>b</sup>
<b><i>Priestia aryabhatai</i> (10<sup>8</sup>CFU/mL) (B)</b>						
Uninoculated	3.74 <sup>b</sup>	10.0 <sup>b</sup>	1.23 <sup>b</sup>	6.99 <sup>b</sup>	1.79 <sup>b</sup>	31.8 <sup>b</sup>
Inoculated	4.25 <sup>a</sup>	12.2 <sup>a</sup>	1.46 <sup>a</sup>	8.04 <sup>a</sup>	2.70 <sup>a</sup>	38.8 <sup>a</sup>
F (A)	**	**	**	**	*	**
F (B)	**	**	**	**	*	**
F (AxB)	ns	ns	ns	ns	ns	ns
CV (%)	23.6	15.5	14.3	17.0	15.8	20.7

The different letters in the same column indicate significant differences at 5% (\*), at 1% (\*\*), and insignificant difference at 5% (ns); DAS: days after sowing; CV: coefficient of variation.

The highest fresh yield of baby corn ear (15.0 t/ha) in the application of 243.8 kg N/ha, the lowest fresh yield of baby corn ear (6.02 t/ha) without N<sub>2</sub> fertilizer application. The fresh yield of baby corn ear ranged from 6.02 to 15.0 t/ha at five different N<sub>2</sub> fertilizer rates. Fresh yield of corn ear (12.2 t/ha) at treatments of *Priestia aryabhatai* inoculum were higher without *Priestia aryabhatai* inoculum (10.0 t/ha). The interaction of N rates × *Priestia aryabhatai* was not significant; all mean values for each treatment are showed in Table 4. Similar, the interaction of N<sub>2</sub> rates × entophytic bacteria were insignificant differences on fresh yields of baby corn silk, husk and edible cob (Table 4). The highest silk yield (2.18 t/ha) was in the application of 325 kg N/ha, the lowest fresh yield of baby corn silk had 0.831 t/ha without N<sub>2</sub> application of 0.0 kg N/ha. The average fresh silk yield (P<0.01) ranged from 0.831 to 2.18 t/ha at N<sub>2</sub> rates, and 1.23 to 1.46 t/ha at *Priestia aryabhatai* inoculum. The average silk yield, which obtained at treatments of *Priestia aryabhatai* inoculum (1.46 t/ha), was higher than without *Priestia aryabhatai* inoculum (1.23 t/ha) and insignificant differences (P < 0.01) on the fresh silk yield of baby corn.

The results in Table 4 showed that there were significant differences among treatments of five N<sub>2</sub> fertilizer rates (p < 0.01) (0, 81.3, 162.5, 243.8 and 325 kg/ha) and *Priestia aryabhatai* (p < 0.01) on fresh husk yield of baby corn. But, the interaction of N rates × Entophytic bacteria was insignificantly different on fresh husk yield of baby corn. Nitrogen ratios (p<0.05), *Priestia aryabhatai* (p<0.05) had significant effect on fresh yield of edible cob. Data on Table 4 showed that highest edible cob yield of baby corn (3.0 t/ha) was recorded at N<sub>2</sub> levels (162.5 kg N/ha), in contrast, application of 0.0 t N/ ha (control) produced lowest cob yield (1.39 t/ha). The fresh cob yield was obtained 2.70 t/ha at treatment of *Priestia aryabhatai* inoculum, which was higher than without *Priestia aryabhatai* inoculation (1.79 t/ha). Nitrogen application significantly improved yield components and yield of plants. The results of research of Yang et al., (2021), fertilizer application was less effect on rhizosphere bacteria, but the significant effect was on

rhizobial strains the legume nodules. This was though the effects of N<sub>2</sub> levels on the soil fertility, and the benefit relation between soil nutrients and the rhizosphere bacteria population. Results of Abdel-Gayed et al., (2019) discovered the entophytic bacterium strain inoculation for crop, which could be affected by soil chemical properties, further, rhizosphere entophytic bacteria could be used as a positive bacterium to increase growth and yield of crops.

**Table 5: effect of N<sub>2</sub> ratios and *Priestia aryabhatai* on baby corn Quality**

Factor	Baby corn cob quality (%)				
	Moisture	Lipid	Protein	Phosphorous	Potassium
<b>Nitrogen rates (kg/ha) (A)</b>					
0.00	82.9 <sup>ab</sup>	0.170 <sup>c</sup>	2.00 <sup>c</sup>	0.045 <sup>c</sup>	0.241 <sup>b</sup>
325	87.6 <sup>a</sup>	0.180 <sup>c</sup>	2.32 <sup>b</sup>	0.066 <sup>b</sup>	0.245 <sup>b</sup>
243.8	86.6 <sup>a</sup>	0.220 <sup>b</sup>	2.62 <sup>a</sup>	0.054 <sup>c</sup>	0.279 <sup>a</sup>
162.5	88.9 <sup>a</sup>	0.130 <sup>d</sup>	2.74 <sup>a</sup>	0.217 <sup>a</sup>	0.250 <sup>b</sup>
81.3	78.3 <sup>b</sup>	0.240 <sup>a</sup>	2.37 <sup>b</sup>	0.052 <sup>c</sup>	0.242 <sup>b</sup>
<b><i>Priestia aryabhatai</i> (10<sup>8</sup>CFU/mL) (B)</b>					
Uninoculated	78.5 <sup>b</sup>	0.174 <sup>b</sup>	2.23 <sup>b</sup>	0.080 <sup>b</sup>	0.233 <sup>b</sup>
Inoculated	91.2 <sup>a</sup>	0.202 <sup>a</sup>	2.59 <sup>a</sup>	0.093 <sup>a</sup>	0.270 <sup>a</sup>
F(A)	**	**	**	**	**
F(B)	**	**	**	**	**
F(A x B)	ns	ns	ns	*	ns
CV (%)	11.7	23.5	15.3	17.5	12.2

The different letters in the same column indicate significant differences at 5% (\*), at 1% (\*\*), and insignificant difference at 5% (ns); DAS: days after sowing; CV: coefficient of variation.

The results revealed that different N<sub>2</sub> rates had a considerable effect on baby corn cob moisture concentration (Table 5). The N<sub>2</sub> fertilizer weight (162.5 kg N/ ha) obtained the highest cob moisture (88.9%), while the N<sub>2</sub> fertilizer weight (81.3 kg N/ha) provided the lowest (78.3%) cob moisture. The average cob moisture of *Priestia aryabhatai* inoculation (P<0.01) valued 78.5% (uninoculated) and 91.2% (inoculated). The cob moisture obtained at treatment of *Priestia aryabhatai* inoculation, which was higher than cob moisture of non *Priestia aryabhatai* inoculation. Their interaction of N rates × *Priestia aryabhatai* was insignificant differences on baby corn cob moisture. The N<sub>2</sub> fertilizer impact of 81.3 kg N/ha recorded the highest lipid content with 0.240% and lowest value of lipid was 0.170% at control treatment and the differences were statistically significant at level 1%. The average lipid concentration of *Priestia aryabhatai* inoculation (P<0.01) valued 0.174% (uninoculated) and 0.202% (inoculated). The lipid value, which obtained at treatment of *Priestia aryabhatai* inoculation, was higher than lipid value of non *Priestia aryabhatai* inoculation. However, their interaction of N rates × *Priestia aryabhatai* was not significant differences on baby corn lipid contents (Table 5). The phosphorous and potassium concentration of baby corn cob was significantly impacted by different levels of different N<sub>2</sub> ratios. The highest phosphorous and potassium values were 0.217% (162.5 kg N/ha) and 0.279 (243.8 kg N/ha), respectively, with statistical differences (p<0.01), and the lowest the phosphorous (0.045%) and potassium (0.241%) value was presented by control treatments (no N<sub>2</sub> application). The average P and K concentration of



*Priestia aryabhatai* inoculation ( $P < 0.01$ ) valued 0.08% (uninoculated) and 0.233% (inoculated), respectively. The P and K content, which observed at treatment of *Priestia aryabhatai* inoculation, was higher than those of non *Priestia aryabhatai* inoculation. Nonetheless, their interaction of N rates  $\times$  *Priestia aryabhatai* was significant differences on baby corn P contents (except K content). According to Eltelib et al. (2006), the relation between pod quality and soil N<sub>2</sub> concentration was significant an effect. Further, the studied results found out where nitrogen remarkably raised the quality composition of baby maize. Thus, recent study has discovered that increased N levels increased baby corn cob contents (Lipid, protein, P and K) in cod maize. Otherwise, low N rate not only restrict pod yield but also cob quality including moisture, lipid, protein, P and K contents (Tsai et al., 1992; Hammad et al., 2011).

## CONCLUSION

This research discovered the impacts of N<sub>2</sub> fertilizer ratios and *Priestia aryabhatai* inoculation on soil fertility, baby corn yield and quality. The discovery revealed that raised fertilizer application had significant effect on either yield and yield attributes or cob quality, such as moisture, oil, protein, P and K contents. Nonetheless, out of the five N<sub>2</sub> ratios applied, the optimal N level between 162.5–243.8 kg N /ha and *Priestia aryabhatai* inoculation might potentially raise the edible cob yield, indicating that N treatment has the ability to produce higher grain yield as well as higher quality contents of baby corn edible cob. Further, all quality and fresh yield properties of baby corn edible cob can be enhanced with the right rate of N fertilizer and *Priestia aryabhatai* inoculation, promoting its better nutritional quality and creating it of greater important agriculture in the future.

## References

- 1) Rani, P.L, Sreenivas, G., & Katti, G.S. (2015). Baby corn based inter cropping system as an alternative pathway for sustainable agriculture. International Journal of Current Microbiology and Applied Sciences. 4(8):869-873.
- 2) Gondaliya, B.R., Desai, K.D., Ahlawat, T.R., Mangroliya, R.M. & Mandaliya, J.V. (2022). Effect of chemicals on growth and yield of baby corn (*Zea mays* L.). The Pharma Innovation Journal. 11(9): 2761-2764.
- 3) Torbert, H.A., Potter, K.N., & Morrison, J.E. Jr (2011) Tillage system, fertilizer nitrogen rate, and timing effect on corn yields in the Texas Blackland Prairie. Agronomy Journal 93:1119-1124.
- 4) Hopf, N., Plesofsky-, N., & Brambl, R. (1992) The heat shock response of pollen and other tissues of maize. Plant Molecular Biology 19, 623-630.
- 5) John, G.W., & Schmitt, M.A. (2007). Advisability of fall-applying nitrogen. Proceedings of the 2008 Wisconsin Fertilizer, Aglime and Pest Management Conference, held on the 15 -17th January, 2008 at University of Wisconsin, Madison, WI. pp. 90-96.
- 6) McCullough, D.E., Girardin, P., Mihajlovic. M., Aguilera, A., & Tollenaar, M. (1994). Influence of N supply on development and dry matter accumulation of an old and new maize hybrid. Canadian Journal of Plant Science. 74, 471-477

- 7) Evans, L.T. (2008) Feeding the Ten Billion. Plants and population growth. Cambridge University Press, p.247
- 8) Subramanian, R.L.Jr, Manonand, S.V. & Rhoads, F.M. (2006) Effect of time, rate, and increment of applied fertilizer on nutrient plant uptake and yield of corn (*Zea mays* L.). Proceedings of Soil Science. 36, 181-184
- 9) Carpici, E.B., Celik, N. & Bayram, G. (2010) Yield and quality of forage maize as influenced by plant density and nitrogen rate. Turkish Journal of Field Crops. 15: 128-132.
- 10) Blackmer, A.M., Pottker, D., Cerrato, M.E. & Webb, J. (2009) Correlations between soil nitrate concentrations in late spring and corn yields in Iowa. American Society of Agronomy. 2, 103-109.
- 11) Bundy, L.G., Andraski, T.W., Ruark, M.D. & Peterson, A.E. (2011). Longterm continuous maize and nitrogen fertilizer effects on productivity and soil properties. Agronomy Journal. 103, 1346-1351.
- 12) Nguyen Van Chuong, Le Minh Tuan, Tran Le Kim Tri, Nguyen Ngoc Phuong Trang, Ho Thanh Tuan, & Nguyen Thi Thuy Diem (2023). Efficient evaluation of *E. asburiae*, *K. quasipneumoniae* and *E. cloacae* combination with vermicompost on growth and yield of groundnut. Seybold report. 18:1862-1870.
- 13) Lau, J.A. & Lennon, J.T. (2011). Evolutionary ecology of plant–microbe interactions: soil microbial structure alters selection on plant traits. New Phytol. 192, 215–224.
- 14) Bever, J.D., Broadhurst, L.M. & Thrall, P.H. (2013). Microbial phylotype composition and diversity predicts plant productivity and plant–soil feedbacks. Ecol. Lett. 16, 167–174.
- 15) Shao, Y., Liu, T., Eisenhauer, N., Zhang, W., Wang, X., Xiong, Y., et al.. (2018). Plants mitigate detrimental nitrogen deposition effects on soil biodiversity. *Soil Biol. Biochem.* 127, 178–186.
- 16) Liu, J., Cui, X., Liu, Z., Guo, Z., Yu, Z., Yao, Q., et al.. (2019). The diversity and geographic distribution of cultivable bacillus-like bacteria across black soils of Northeast China. *Front. Microbiol.* 10:1424.
- 17) Roy, K., Ghosh, D., DeBruyn, J. M., Dasgupta, T., Wommack, K. E., Liang, X., et al.. (2020). Temporal dynamics of soil virus and bacterial populations in agricultural and early plant successional soils. *Front. Microbiol.* 11:1494.
- 18) Nguyen Van Chuong. (2023). Response of peanut quality and yield to chicken manure combined with Rhizobium inoculation in sandy soil. *Communications in Science and Technology* 8(1) (2023) 31–37.
- 19) Korenblum, E., Dong, Y., Szymanski, J., Panda, S., Jozwiak, A., Massalha, H., et al.. (2020). Rhizosphere microbiome mediates systemic root metabolite exudation by root-to-root signaling. *Proc. Natl. Acad. Sci. U. S. A.* 117, 3874–3883.
- 20) Berendsen, R.L., Pieterse, C.M., & Bakker, P.A. (2012). The rhizosphere microbiome and plant health. *Trends Plant Sci.* 17, 478–486.
- 21) Mendes, R., Garbeva, P., & Raaijmakers, J. M. (2013). The rhizosphere microbiome: significance of plant beneficial, plant pathogenic, and human pathogenic microorganisms. *FEMS Microbiol. Rev.* 37, 634–663.
- 22) Lazcano, C., Boyd, E., Holme, G., Hewavitharana, S., Pasulka, A. & Ivors, K. (2021). The rhizosphere microbiome plays a role in the resistance to soil-borne pathogens and nutrient uptake of strawberry cultivars under field conditions. *Sci. Rep.* 11:3188.
- 23) Moreau, D., Bardgett, R.D., Finlay, R.D., Jones, D.L. & Philippot, L. (2019). A plant perspective on nitrogen cycling in the rhizosphere. *Funct. Ecol.* 33, 540–552.
- 24) Korenblum, E., Dong, Y., Szymanski, J., Panda, S., Jozwiak, A., Massalha, H., et al.. (2020). Rhizosphere microbiome mediates systemic root metabolite exudation by root-to-root signaling. *Proc. Natl. Acad. Sci. U. S. A.* 117, 3874–3883.

- 25) Ahmed, A. & Hasnain S. (2014). Auxins as one of the factors of plant growth improvement by plant growth promoting rhizobacteria. *Pol. J. Microbiol.* 63, 261–266.
- 26) Chen, X.H., Koumoutsis, A., Scholz, R., Eisenreich, A., Schneider, K., Heinemeyer, I., et al. (2007). Comparative analysis of the complete genome sequence of the plant growth-promoting bacterium *Bacillus amyloliquefaciens* FZB42. *Nat. Biotechnol.* 25, 1007–1014.
- 27) Backer, R., Rokem, J. S., Ilangumaran, G., Lamont, J., Praslickova, D., Ricci, E., et al. (2018). Plant growth-promoting rhizobacteria: context, mechanisms of action, and roadmap to commercialization of biostimulants for sustainable agriculture. *Front. Plant Sci.* 9:1473.
- 28) Tahir, H.A., Gu, Q., Wu, H., Raza, W., Hanif, A., Wu, L., et al. (2017). Plant growth promotion by volatile organic compounds produced by *Bacillus subtilis* SYST2. *Front. Microbiol.* 8:171.
- 29) Shivaji, S., Chaturvedi, P., Begum, Z., Pindi, P.K., Manorama, R., Padmanaban, D.A., et al. (2009). *Janibacter hoylei* sp. nov., *Bacillus isronensis* sp. nov. and *Bacillus aryabhatai* sp. nov., isolated from cryotubes used for collecting air from the upper atmosphere. *Int. J. Syst. Evol. Microbiol.* 59, 2977–2986.
- 30) Bhattacharyya, C., Bakshi, U., Mallick, I., Mukherji, S., Bera, B. & Ghosh, A. (2017). Genome-guided insights into the plant growth promotion capabilities of the physiologically versatile *Bacillus aryabhatai* strain AB211. *Front. Microbiol.* 8:411.
- 31) Park, Y.G., Mun, B.G., Kang, S.M., Hussain, A., Shahzad, R., Seo, C.W., et al. (2017). *Bacillus aryabhatai* SRB02 tolerates oxidative and nitrosative stress and promotes the growth of soybean by modulating the production of phytohormones. *PLoS One* 12: e0173203.
- 32) Ghosh, P.K., Maiti, T.K., Pramanik, K., Ghosh, S.K., Mitra, S. & De, T. K. (2018). The role of arsenic resistant *Bacillus aryabhatai* MCC3374 in promotion of rice seedlings growth and alleviation of arsenic phytotoxicity. *Chemosphere* 211, 407–419.
- 33) Mehmood, S., Khan, A.A., Shi, F., Tahir, M., Sultan, T., Munis, M.F.H., et al. (2021). Alleviation of salt stress in wheat seedlings via multifunctional *Bacillus aryabhatai* PM34: an in-vitro study. *Sustainability* 13, 8030–8030.
- 34) Chu, T.N., Bui, L.V. & Hoang, M.T. (2020). *Pseudomonas* PS01 isolated from maize rhizosphere alters root system architecture and promotes plant growth. *Microorganisms* 8:471.
- 35) Chen, Y.F., Yang, H., Shen, Z.Z. & Ye, J.R. (2022). Whole-genome sequencing and potassium-solubilizing mechanism of *Bacillus aryabhatai* SK1-7. *Front. Microbiol.* 12:722379
- 36) Deng, C., Zhang, N., Liang, X., Huang, T. & Li B. (2022). *Bacillus aryabhatai* LAD impacts rhizosphere bacterial community structure and promotes maize plant growth. *J. Sci. Food Agric.* 102, 6650–6657.
- 37) Chuong, N.V., Trang, N.N.P, Tri, T.L.K.T., Loan, L.T.T.L., Son, N.T.T.S., Tuan, H.T. & Cuong, T.V. (2023). Isolation and Genomic Identification of Rhizosphere N<sub>2</sub>-Fixing Bacteria from groundnut nodules. *Daxue Xuebao (Gongxueban)/Journal of Jilin University (Engineering and Technology Edition)*. 42, 141-151.
- 38) Carterand, M.R. & Gregorich,E.G. (2007). Soil sampling and methods of analysis. Second Edition, Virgil: Georgics (II, 490).
- 39) Ren, T.; Li, H.; Lu, J.; Bu, R.; Li, X.; Cong, R. & Lu, M. (2015). Crop rotation-dependent yield responses to fertilization in winter oilseed rape (*Brassica napus* L.). *Crop J.* 3, 396–404.
- 40) Peng, N., Oladele, O., Song, X., Ju, X., Jia, Z., H Hu, et al. (2022). Opportunities and approaches for manipulating soil-plant microbiomes for effective crop nitrogen use in agroecosystems. *Front. Agric. Sci. Eng.* 2022; 9, 333–343.
- 41) Fan, K., Delgado-Baquerizo, M., Guo, X., Wang, D., Wu, Y., Zhu, M., Yu, W., Yao, H., Zhu, Y.G. & Chu, H. (2019). Suppressed N fixation and diazotrophs after four decades of fertilization. *Microbiome.* 7, 143.

- 42) Zhang, R., Mu, Y., Li, X., Li, S., Sang, P., Wang, X., Wu, H., Xu, N. (2020). Response of the arbuscular mycorrhizal fungi diversity and community in maize and soybean rhizosphere soil and roots to intercropping systems with different nitrogen application rates. *Sci. Total Environ.* 740, 139810.
- 43) Xia, Z., Yang, J., Sang, C., Wang, X., Sun, L., Jiang, P., et al. (2020). Phosphorus reduces negative effects of nitrogen addition on soil microbial communities and functions. *Microorganisms* 8 (11), 1828.
- 44) Ran, J., Liu, X., Hui, X., Ma, Q., & Liu J. (2021). Differentiating bacterial community responses to long-term phosphorus fertilization in wheat bulk and rhizosphere soils on the loess plateau. *Appl. Soil Ecol.* 166-172.
- 45) Leu, A.F. (2005). Organic lychee and rambutan production. *Acta Hort.* 665, 241-248.
- 46) Monaco, S., Hatchb, D.J., Saccoa, D., Bertoraa, C., & Grignania, C. (2008). Changes in chemical and biochemical soil properties induced by 11-yr repeated additions of different organic materials in maize-based forage systems. *Soil Biology & Biochemistry.* 40(3), 608-615.
- 47) Bengtson, P., Barker, J., & Grayston, S.T. (2012). Evidence of a strong coupling between root exudation, C and N availability, and stimulated SOM decomposition caused by rhizosphere priming effects. *Ecology and Evolution.* 2, 1843– 1852.
- 48) Bade, N.E., & Cheng, W.X. (2007). Rhizosphere priming effect of *Populus fremontii* obscures the temperature sensitivity of soil organic carbon respiration. *Soil Biology & Biochemistry.* 39, 600– 606.
- 49) Moosavi, S.G., Seghatoleslami, M.J. & Moazeni, A. (2012). Effect of planting date and plant density on morphological traits, LAI and forage corn (*Sc. 370*) yield in second cultivation. *International Research Journal of Applied and Basic Sciences* 3: 57-63
- 50) Thavaprakaash, N. & Velayudham, K. (2009). Influence of crop geometry, intercropping system and INM practices on productivity of baby corn (*Zea mays L.*) based intercropping. *Mysore Journal of Agricultural Sciences* 43: 686 – 695.
- 51) Kar, P.P., Barik, K.C., Mahapatra, P.K., Garnayak, L.M., Rath, B.S., Bastia, D.K. & Khanda, C.M. (2006). Effect of planting geometry and nitrogen on yield, economics and nitrogen uptake of sweet corn. *Indian Journal of Agronomy* 51: 43-53.
- 52) Eltelib, H.A., Hamad, M.A. & Ali, E.E. (2006). The effect of nitrogen and phosphorus fertilization on growth, yield, and quality of forage maize (*Zea mays L.*). *Journal of Agronomy*, 5(3): 515–518.
- 53) Tsai, C.Y., Dweikat, I., Huber, D.M. & Warren, H.L. (1992). Interrelationship of nitrogen nutrition with maize (*Zea mays*) grain yield, nitrogen use efficiency and grain quality. *Journal of the Science of Food and Agriculture*, 58(1): 1–8.
- 54) Hammad, H.M., Ahmad, A., Khaliq, T., Farhad, W., and Mubeen, M. (2011). Optimizing rate of nitrogen application for higher yield and quality in maize under semiarid environment. *Crop and Environment*, 2(1): 38–41.
- 55) Yang, Z., Xiao, Z.S., Chen, S., Liu, J., Zhu, W., Xu, Q., Li, L., Guo, F. & Lan, L. (2021). Effect of nitrogen application rates on the yield and quality of different oleic peanuts. *Journal of Henan Agricultural Sciences.* 50(9), 44–52.
- 56) Abdel-Gayed, M., Ahmad, Abo-Zaid, G., Attia, Matar, S., Mohamed, Hafez, E., & Elsayed. (2019). Fermentation, formulation and evaluation of PGPR *Bacillus subtilis* isolate as a bio agent for reducing occurrence of peanut soil-borne diseases. *Journal of Integrative Agriculture*, 18(9), 2080-2092